BIOCHEMISTRY AND MOLECULAR BIOLOGY (BMB)

BMB 1: Understanding the Bases of Human Disease

3 Credits

A broad survey of the molecular and cellular factors that contribute to an understanding of selected human diseases. B M B 001 Understanding the Bases of Human Disease (3) (GN)(BA) This course meets the Bachelor of Arts degree requirements. B M B 001, Understanding the Bases of Human Disease, is a survey of the most common diseases of humans. In addition to discussing various diseases, symptoms, outcomes, prevention and treatment options, we also study disease processes by describing events at the cellular or DNA level. We will come to appreciate that various cellular, metabolic or genetic problems can give rise to disorders that carry a common name - anemia, for example. One form, Sickle Cell anemia, also provides a good example of another theme of the course: how a small genetically-based problem in one function of one cell type can lead to a cascade of malfunctions with ramifications system wide. How medicines work and how they are metabolized by the body will be discussed. Treatment of the growing problem of drug/drug, drug/food, and drug/supplement interactions will also be included. Any study of infectious disease must be prefaced by a discussion of the proper functioning of the immune system and how its various components work together to fight an infection. The proper function or malfunction of the immune system is also studied for relationships to noninfectious diseases such as cancer and certain genetic disorders. As with any system of the body, the immune system also has its own unique set of disorders: allergy, hypersensitivity, autoimmunity and immune deficiency - congenital and acquired. In our exploration of infectious diseases, we will look at various mechanisms of infection used by different types of organisms and at strategies the offending organisms might have to fool the immune system, hide from the immune system, or fight against it. A prime example is the ability of HIV to hide from the immune system. Knowing disease mechanisms and the life cycle of the infectious agent can suggest strategies for treatment of the disease. Another focus of this unit will include discussion of emerging and re-emerging diseases. Human genetic traits and susceptibilities are discussed throughout the course, but we will also address the major genetic disorders, patterns of inheritance, chromosomal disorders and new treatments available for a growing number of such disorders. Included under this topic is the problem of cancer in all of its various forms today. The course addresses issues of normal cell/cancer cell characteristics and the concept of cancer as a "genetic disease." Using the information gleaned from units on cell function, genetics and the immune system, we will explore new treatment options evolving today. No course on treatment of disease is complete without the study of the two major "lifestyle" diseases: coronary vascular disease and diabetes. We will explore the environmental factors as well as the genetic and infection-related factors often associated with these diseases. The course is independent of all other B M B courses and does not serve as a prerequisite for any other course.

Bachelor of Arts: Natural Sciences
General Education: Natural Sciences (GN)
BMB 212: Elementary Biochemistry Laboratory

1 Credits/Maximum of 1

Selected experiments to illustrate major biochemical principles and techniques. B M B 212 B M B 212 Elementary Biochemistry Laboratory (1)B M B 212 exposes students to techniques typically used in industrial and academic laboratories to isolate proteins, perform enzyme kinetics, characterize carbohydrates and lipids, and study molecular biology. Data interpretation and conceptual understanding are emphasized. Specifically, students determine a weak acid’s buffer range with a pH meter; quantify protein concentrations using a spectrophotometer; partially purify acid phosphatase from wheat germ, using ammonium sulfate precipitation, centrifugation and dialysis; characterize acid phosphatase kinetics; subject glycogen to acid and salivary amylase hydrolysis, then compare products using thin layer chromatography; isolate plasmid DNA from E.coli, then digest the plasmid DNA with restriction enzymes and analyze the products using agarose gel electrophoresis; make soap from commercial oils and fats; and lastly, identify fatty acids using a gas chromatograph. Students write laboratory reports to present their findings and correlate theory with actual experimental results. Written quizzes assess conceptual understanding of experiments. Teaching assistant evaluations judge the student’s level of laboratory skill achievement, preparation, and ability to work with others in a professional manner. A solid chemistry knowledge base (CHEM 012 and either CHEM 034 or 038), previous laboratory experience, and the ability to work with mathematical word problems are expected of all students enrolled in the course. Biochemistry focuses on the chemistry of living organisms. This course provides basic biochemistry laboratory skills and exposure to widely-used methodology to develop a fundamental understanding of biochemistry needed for advanced courses in the student’s major and compatible with the student’s career interests. Outside resources for the student include reserved books and a course web site: www.bmb.psu.edu/courses/daniel/BMB212/default.htm.

Enforced Prerequisite: or concurrent: BMB 211

BMB 221: Applied Biochemistry

2 Credits

Application and correlation of biochemical events to physiological-nutritional processes in specialized cells, fluids, and whole animals. Students may not receive credit for both B M B 221 and 401. B M B 221 B M B 221 Applied Biochemistry (2)In B M B 221, students will build on their knowledge from Elementary Biochemistry (B M B 211). Emphasis is on the application of biochemical principles to specific problems in medicine, agriculture, pharmaceuticals and biotechnology. Students review fundamental principles and learn how biochemical methods, techniques and theory are applied. Class presentations are computer based in some cases. The course may also include class discussions. Evaluation and grading varies with the instructor, but possible methods of evaluation include multiple choice examinations, essay-short answer questions, and in-class contributions. Students will have completed at least three semesters of chemistry before B M B 221, because Elementary Biochemistry (B M B 211) is a prerequisite, and inorganic (CHEM 012) and organic chemistry (CHEM 034 or CHEM 038) are prerequisites for B M B 211. Knowledge of organic chemistry and basic biochemistry is essential so that the course can focus on applications rather than introductory material. B M B 221 is a requirement for those Biotechnology majors who choose not to take the 400-level series of introductory biochemistry courses. It is also required of students enrolled in the science option of the Dairy and Animal Science major. It serves as an elective for all other majors. It is not approved as a general education course.

Enforced Prerequisite: BMB 211

BMB 251: Molecular and Cell Biology I

3 Credits

BMB 251 Molecular and Cell Biology I (3) is an introduction to the fundamental principles of molecular and cellular biology, with a primary focus on eukaryotic cells. Topics covered will include elementary biochemistry; structure and function of biological macromolecules, the cell and its organelles; the role of biological membranes in bioenergetics and sub-cellular compartments. There will be a particular emphasis on the molecular mechanism of heredity; the organization and expression of genetic information; experimental methods used in the analysis of gene expression and the relationship between gene/protein structure and function.

Enforced Prerequisite: CHEM 112 or CHEM 112H Recommended preparations MICRB 201 or MICRB 201H Cross-listed with: MICRB 251

BMB 251H: Molecular and Cell Biology I

3 Credits

Biomolecules, genetic mechanisms, organization of cells and their organelles, DNA replication, protein synthesis, membranes, the cell nucleus, energy conversion. BMB 251H Molecular and Cell Biology I (3) This course is an introduction to the fundamental principles of molecular and cellular biology, with a primary focus on eukaryotic cells. Topics covered will include elementary biochemistry; structure and function of biological macromolecules, the cell and its organelles; the role of biological membranes in bioenergetics and sub-cellular compartments. There will be a particular emphasis on the molecular mechanism of heredity; the organization and expression of genetic information; experimental methods used in the analysis of gene expression and the relationship between gene/protein structure and function. A key feature of the Honors course is the use of review papers and peer-reviewed journal articles as integral components of the course. The objectives of this component of the Honors course are to: 1) introduce students to the scientific method (the formulation of hypotheses based on observation and the processes underpinning the rigorous test of such hypotheses); and 2) provide the intellectual framework for a critical evaluation of the literature. Students are expected to engage in classroom discussion and will be evaluated by a combination of classroom presentations, multiple choice and short essay exams. Students are expected to develop a "big picture" view of how the various cellular processes are related to each other and also attain a thorough understanding of the molecular details of the individual processes (e.g. the order and molecular details of events leading from transcription to protein localization within a cell).

Enforced Prerequisite: CHEM 112 or CHEM 112H Honors

BMB 252: Molecular and Cell Biology II

3 Credits

Continuation of BMB 251 / MICRB 251; cytoskeleton, cell growth, division, adhesion, signalling, germ cells, differentiation, immune system, nervous system, plant cells. BMB 252 / MICRB 252 Molecular and Cell Biology II (3) focuses on the internal organization on eukaryotic
cells and their organization in multi-cellular organisms. Topics covered include cell communication, the cytoskeleton, cell cycle, fertilization and development of multi-cellular organisms, genesis of tissues, and the molecular mechanisms of cancer and immunity.

**Enforced Prerequisite:** BMB 251 or MICRB 251 or BMB 251H or BIOL 230W or BIOL 230M

**Cross-listed with:** MICRB 252

**BMB 252H: Molecular and Cell Biology II**

3 Credits

Continuation of BMB 251H; cytoskeleton, cell growth, division, adhesion, signaling, germ cells, differentiation, immune system, nervous system, plant cells. BMB 252H Molecular and Cell Biology II (3) This course is the second part of BMB 251H. This section focuses on the internal organization on eukaryotic cells and their organization in multi-cellular organisms. Topics covered include cell communication, the cytoskeleton, cell cycle, fertilization and development of multi-cellular organisms, genesis of tissues, and the molecular mechanisms of cancer and immunity. There will be a particular emphasis on how the basic principles and experimental approaches presented in BMB 251H are employed to address questions related to the topics that will be covered in BMB 252H. As in BMB 251H, a key feature of the Honors section will be the use of review papers and peer-reviewed journal articles as integral components of the course. The objectives of this component of the Honors section are to: 1) introduce students to the scientific method (the formulation of hypotheses based on observation and the processes underpinning the rigorous test of such hypotheses); and 2) provide the intellectual framework for a critical evaluation of the literature. Students are expected to engage in classroom discussion and will be evaluated by a combination of classroom presentations, multiple choice and short essay exams.

**Enforced Prerequisite:** BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M Recommended preparations BMB 251H or BIOL 230M

**Honors**

**BMB 294: Research Projects**

1-6 Credits/Maximum of 8

Supervised student activities on research projects identified on an individual or small-group basis.

**BMB 398: Special Topics**

1-9 Credits/Maximum of 9

Formal courses given infrequently to explore, in depth, a comparatively narrow subject that may be topical or of special interest.

**BMB 399: Foreign Studies**

1-12 Credits/Maximum of 12

Courses offered in foreign countries by individual or group instruction.

**International Cultures (IL)**

**BMB 400: Molecular Biology of the Gene**

2-3 Credits

Biochemistry of genetic phenomena, including the structure, replication and dynamics of genes and chromosomes, their expression and regulation. BMB 400 Molecular Biology of the Gene (2-3) Molecular Biology of the Gene examines the flow of information in living things at the molecular level. Topics such as the following are included: 1) DNA replication, repair and recombination, 2) RNA transcription and modification, and 3) protein translation, folding and modification. This class is designed as a one-semester course having the objectives of understanding concepts in molecular biology and gene regulation, and exploring research materials and methods used in the laboratory. Course materials are prepared not only from the textbook but also from the primary literature. Therefore, students who want to take this course should have some familiarity with reading research articles. BMB 400 is for advanced undergraduates who have already taken introductory molecular biology and biochemistry. Knowledge of molecular biology is essential background for pursuit of a career in the life sciences, including academia, medicine, industry, forensic science and science policy.

**Enforced Prerequisite:** (BMB 251 OR MICRB 251 OR BIOL 230W OR BMB 251H OR BIOL 230M ) AND ( CHEM 212 OR CHEM 212H )

**BMB 400A: Molecular Biology of the Gene**

3 Credits/Maximum of 3

Molecular Biology of the Gene examines the flow of information in living things at the molecular level. Topics such as the following are included: 1) DNA replication, mutation, repair and recombination, 2) RNA transcription and modification, 3) protein translation, folding and modification, and 4) gene regulation. This class is designed as a one-semester course having the objectives of understanding concepts in molecular biology and gene regulation, and exploring research materials and methods used in the laboratory. Subjects are related to forensic applications when applicable. Course materials are prepared not only from the textbook but also from the primary literature. Therefore, students who want to take this course should have some familiarity with reading research articles.

**Enforced Prerequisites:** ( BIOL 222 or BIOL 322 ) AND ( BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M ) AND ( CHEM 212 or CHEM 212H )

**BMB 401: General Biochemistry**

3 Credits

Principles of the structure and function of biological molecules, including carbohydrates, lipids, membranes, proteins, and enzymes. BMB 401 is the first course of the general biochemistry series, a sequence designed to prepare students for careers and graduate study in the life sciences. Overall, biochemistry describes, in chemical and molecular terms, the structures, mechanisms, and chemical processes at work in all living things, and abstracts organizing principles that underlie life in all its diverse forms. Building upon concepts introduced in molecular and cellular biology and in organic chemistry, students in BMB 401 synthesize and apply this knowledge toward understanding the structure and function of the major classes of cellular constituents: water, and the various macromolecules – amino acids and proteins, sugars, and polysaccharides, nucleotides and nucleic acids, fatty acids and lipids, and membranes and various membrane proteins. These molecules interact...
to comprise the next level of multi-and mixed molecular structures and organelles that enable a cell to carry out its many metabolic functions. Students also learn about the technologies used to study cellular components and processes, and current advances in biotechnology that have accelerated the pace of discovery in the field. Having gained familiarity with the molecules found in a cell, students are well-equipped to take on more advanced topics in the exciting, rapidly-evolving fields of the life sciences. An overriding theme in biochemistry is that polymers of living systems, though structurally large and functionally complex, are highly ordered chemical entities, with specific sequences of monomeric subunits giving rise to discrete structures and functions. The course includes all of the following topics: 1) an introduction to protein structure function relationships, covering the structural basis of protein functions; 2) enzyme kinetics and mechanisms; 3) simple and complex carbohydrates and topics in glycobiology that include energy storage, framework skeleton, and specific molecular recognition; 4) various classes of lipids, including phospholipids, complex lipids, membrane biology and transport systems, and 5) an analysis of the biochemical basis of signal transduction describing how specific signals regulate biomolecular activity both within a cell and between cells to keep an organism in homeostasis. Lastly, as a transition to intermediary metabolism, a bioenergetic framework is introduced to provide a framework for understanding pathways of carbon and nitrogen metabolism, using glycolytic reactions as an example. Students may not receive credit for both BMB 401 and CHEM 476 or BMB 401 and BMB 221.

**Enforced Prerequisites:** (CHEM 210 or CHEM 210H ) AND ( BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M or BME 201)

**Recommended preparations:** CHEM 212 OR CHEM 212H

**BMB 401H: General Biochemistry**

3 Credits

Principles of the structure and function of biological molecules, including carbohydrates, lipids, membranes, proteins, and enzymes. BMB 401H General Biochemistry (3) is the first course of the honors general biochemistry series, a sequence designed to prepare students for careers and graduate study in the life sciences. Overall, biochemistry describes, in chemical and molecular terms, the structures, mechanisms, and chemical processes at work in all living things, and abstracts organizing principles that underlie life in all its diverse forms. Building upon concepts introduced in molecular and cellular biology and in organic chemistry, students in BMB 401H synthesize and apply this knowledge toward understanding the structure and function of the major classes of cellular constituents: water, and the various macromolecules -- amino acids and proteins, sugars, and polysaccharides, nucleotides and nucleic acids, fatty acids and lipids, and membranes and various membrane proteins. These molecules interact to comprise the next level of multi-and mixed molecular structures and organelles that enable a cell to carry out its many metabolic functions. Students also learn about the technologies used to study cellular components and processes, and current advances in biotechnology that have accelerated the pace of discovery in the field. Having gained familiarity with the molecules found in a cell, students are well-equipped to take on more advanced topics in the exciting, rapidly-evolving fields of the life sciences. An overriding theme in biochemistry is that polymers of living systems, though structurally large and functionally complex, are highly ordered chemical entities, with specific sequences of monomeric subunits giving rise to discrete structures and functions. The course begins with an introduction to proteins, covering the structural basis of protein functions and then moves on to enzyme kinetics and mechanisms. Next, students explore simple and complex carbohydrates and topics in glycobiology that include energy storage, framework skeleton, and specific molecular recognition. Various classes of lipids, including phospholipids, complex lipids, membrane biology and transport systems, are covered next. Following is an analysis of the biochemical basis of signal transduction describing how specific signals regulate biomolecular activity within a cell, and between cells-to keep an organism in homeostasis. Lastly, as a transition to intermediary metabolism in BMB 402H, an introduction to bioenergetic principles is included to provide a framework for understanding pathways of carbon and nitrogen metabolism, using glycolytic reactions as an example. Students may not receive credit for both CHEM 476 and BMB 401H.

**Enforced Prerequisites:** (CHEM 212 or CHEM 212H ) and ( BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M or BME 201)

**Honors**

**BMB 402: General Biochemistry**

3 Credits

BMB 402 General Biochemistry (3) is a comprehensive survey of the pathways and regulation of intermediary metabolism. Most, if not all, of the students taking BMB 402 intend to pursue a Ph.D., M.D. or M.D./Ph.D. degree after graduation. Since biochemistry is an important discipline for advanced studies in life and medical sciences, a major goal of BMB 402 is to prepare students well for their future challenges in graduate or medical school. The major topics covered include glycolysis, TCA cycle, metabolism of fatty acids, lipids (phospholipids, cholesterol and sphingolipids), amino acids and nucleotides, signal transduction, and human genetic diseases. Since biochemistry is a very rapidly progressing discipline, any new developments not covered by the textbook are introduced in the lectures or via discussion of current scientific papers at an appropriate level. In addition, students must be able to integrate information learned from different but related material.

**Enforced Prerequisites:** BMB 401 or BMB 401H or CHEM 476

**BMB 402H: General Biochemistry**

3 Credits

Comprehensive survey of the pathways and regulation of intermediary metabolism. BMB 402H General Biochemistry (3) provides a more intimate and interactive class environment than does BMB 402. Most, if not all, of the students taking BMB 402H intend to pursue a Ph.D., M.D. or M.D./Ph.D. degree after graduation. Since biochemistry is an important discipline for advanced studies in life and medical sciences, a major goal of BMB 402H is to prepare students well for their future challenges in graduate or medical school. The textbook used is more advanced than that used in BMB 402, and it is what is typically used in biochemistry courses taught at premier medical schools and graduate schools. The class is conducted at a challenging level to provide students with the opportunity to treat topics in greater depth and to explore current development more fully than is possible in BMB 402. The major topics covered include glycolysis, TCA cycle, metabolism of fatty acids, lipids (phospholipids, cholesterol and sphingolipids), amino acids and nucleotides, signal transduction, and human genetic diseases. Since biochemistry is a very rapidly progressing discipline, any new developments not covered by the textbook are introduced in the lectures or via discussion of current scientific papers at an appropriate level. BMB 402H exams require that students understand all aspects of a particular metabolic pathway under study, including the sequential steps of the pathway, chemical structures of all intermediates, the mechanisms of all key reactions, regulation of the pathway, and the relationships
with other pathways. In addition, students must be able to integrate information learned from different but related material.

**Enforced Prerequisites:** BMB 401 or BMB 401H

**Honors**

BMB 403: Biochemistry Laboratory

1 Credits

An introduction to techniques of experimental biochemistry, illustrating principles covered in BMB 402.

**Enforced Prerequisite:** or concurrent: BMB402

BMB 406: Molecular Biology

3 Credits

A discussion of current aspects of cell molecular biology with a laboratory emphasizing current biotechnology techniques.

**Prerequisite:** BIOL 222 or BIOL 322 ; BIOL 230W or B M B 251 ; CHEM 212

BMB 408: Instructional Practice

1-2 Credits/Maximum of 2

Participation in the instruction of undergraduate laboratory and lecture courses, including classroom preparation; discussion of principles and objectives of each exercise. B M B 408 Instructional Practice (1-2) Students in this course will gain experience in science teaching by participating in either lecture courses (as Learning Assistants) or laboratory courses (as Teaching Assistants) with the goal of making students in this course more effective as teachers and communicators in their future careers in science. Participation in instruction of selected departmental lecture and laboratory courses will include preparation of class materials, and facilitating discussion and presentation of course principles and learning objectives of each topic or exercise.

**Prerequisite:** 10 credits in biochemistry and molecular biology and permission of the department

BMB 411: Survey of Biochemistry and Molecular Biology Literature

1 Credits

An introduction to readings and oral presentations in biochemistry and molecular biology.

**Enforced Prerequisites:** ( BMB 401 or BMB 401H ) and ( BMB 400 or BMB 402 or BMB 402H)

BMB 428: Physical Chemistry with Biological Applications

3 Credits

This course uses the study of physical chemistry to develop quantitative and analytical skills in biological and chemical systems. The course covers thermodynamics, solution chemistry and kinetics. The study of thermodynamics includes the three laws of thermodynamics, and the concepts of energy, entropy and enthalpy. Solution chemistry includes the study of colligative properties, ligand binding and energetics in biological systems. Kinetics covers the study of both chemical and enzyme kinetics, as well as enzyme inhibition.

**Enforced Prerequisites:** ( MATH 141 or MATH 141B or MATH 141H ) and ( PHYS 212 or PHYS 212H or PHYS 251 ) and ( CHEM 202 or CHEM 202H or CHEM 210 or CHEM 210H )

BMB 428H: Physical Chemistry with Biological Applications

3 Credits

Chemical thermodynamics and kinetics with applications to biological problems.

**Honors**

BMB 430: Developmental Biology

3 Credits

Developmental Biology introduces students to one of the most complex and exciting areas of modern biology. This course will cover basic concepts using a comparative embryology approach and focus on molecular and genetic analysis of mechanisms involved in cell differentiation and organ development. Students will learn how studies of development in different model organisms including invertebrate and vertebrate species have identified a network of evolutionarily conserved genes and signaling pathways that regulate embryonic development and morphogenesis.

**Enforced Prerequisites:** BMB 252 or MICRB 252 or BMB 252H or BIOL 230W or BIOL 230M Recommended preparations BIOL 222 OR BIOL 322

Cross-listed with: BIOL 430

BMB 432: Advanced Immunology: Signaling in the Immune System

3 Credits

The study of signaling pathways that regulate the immune response. BMB 432 / MICRB 432 / VBSC 432 Advanced Immunology: Signaling in the Immune System (3)This course will use the immune system as a model in which to study how cells communicate in order to coordinate an immune response. We will focus on signaling mechanisms that regulate such immune responses as T cell activation, Th1/Th2 differentiation, macrophage activation, and migration of immune cells to sites of inflammation. All lectures are based on recent reviews by key investigators in each field, as well as primary articles to present students with the most recent advances, techniques, and approaches used. The goal of the course will be to convey a basic understanding of intracellular signaling mechanisms that will pertain to all areas of biology, an appreciation for current questions and future directions in the field, and an in depth understanding of the signals that govern immune responses. The material presented will build on the basic concepts learned in BMB 400 and MICRB 410, and will lay the foundation for more advanced courses at the graduate level.

**Enforced Prerequisites:** (BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M) and (MICRB 410 or VBSC 410)

Cross-listed with: MICRB 432, VBSC 432

BMB 433: Molecular and Cellular Toxicology

3 Credits

In-depth coverage of processes by which drugs/chemicals interact with biological systems and the experimental approaches used to study these interactions.
Methods. Specifically, the experiments include preparation of buffers to protein biochemistry topics and laboratory techniques typically portion of BMB 442 / MICRB 442 is designed to introduce students the lac operon, a classic molecular genetic model system. The proteins is isolated and analyzed by agarose gel electrophoresis, completing media containing drugs. From the transformed cells, recombinant DNA molecules are identified by selective plating on growth into E. coli cells that initially have no drug resistance. Cells that acquire composition from either original molecule. The recombinant molecules, with the enzyme Ligase, which randomly combines small fragments molecules into smaller fragments. The fragments are mixed and treated into two sources. Students use restriction enzymes to cut two distinct DNA a gene, i.e. to make a recombinant molecule comprised of DNA from. The central experiment entails all basic procedures necessary to clone reinforcement of principles of Molecular Genetics from lecture courses.

Enforced Prerequisites: (MICRB 201 or MICRB 201H) and (BMB 252 or MICRB 252 or BMB 252H) or (BIOL 230W or BIOL 230M)
Cross-listed with: MICRB 435, VBSC 435

BMB 437: Physiological Biochemistry
2 Credits

Physiological aspects of biochemistry, with emphasis on mammalian metabolism, specialized tissue and fluid functions, detoxification mechanisms, energetics, and physiological interrelationships.

Enforced Prerequisite: BMB 402

BMB 442: Laboratory in Proteins, Nucleic Acids, and Molecular Cloning
3 Credits

Laboratory in enzyme purifications and assay techniques; nucleic acid isolation and characterization, including plasmid preparation. BMB 442 / MICRB 442 Laboratory in Proteins, Nucleic Acids, and Molecular Cloning (3) The DNA portion of B M B/MICRB 442 serves as an introduction to fundamental techniques of recombinant DNA technology and as a reinforcement of principles of Molecular Genetics from lecture courses. The central experiment entails all basic procedures necessary to clone a gene, i.e. to make a recombinant molecule comprised of DNA from two sources. Students use restriction enzymes to cut two distinct DNA molecules into smaller fragments. The fragments are mixed and treated with the enzyme Ligase, which randomly combines small fragments into large recombinant DNA molecules in new combinations different in composition from either original molecule. The recombinant molecules, which include genes that confer drug resistance, are transformed into E. coli cells that initially have no drug resistance. Cells that acquire recombinant DNA molecules are identified by selective plating on growth media containing drugs. From the transformed cells, recombinant DNA is isolated and analyzed by agarose gel electrophoresis, completing the array of basic gene cloning techniques. In addition to this central, multi-session experiment, students also do PCR and an investigation of the lac operon, a classic molecular genetic model system. The proteins portion of BMB 442 / MICRB 442 is designed to introduce students to protein biochemistry topics and laboratory techniques typically encountered in academic and commercial settings. Students will learn about buffers, spectroscopy, enzyme purification and characterization methods. Specifically, the experiments include preparation of buffers and performing kinetic studies to determine Km and Vmax values. Separation of a mixture of phycobiliproteins using ion-exchange column chromatography is a major experiment that the students will perform to learn protein purification methods. In this experiment they will learn how to pour a column, apply sample, elute it with salt gradient and collect fractions using automated fraction collector. Ammonium sulfate precipitation and dialysis will be part of protein purification procedures. Characterization of the separated proteins will be performed by determining the absorption spectra with spectrophotometer and by determining the molecular weights of the subunits of the phycobiliproteins by SDS-polyacrylamide gel electrophoresis.

Enforced Prerequisites: (CHEM 202 or CHEM 202H or CHEM 210 or CHEM 210H ) and (BMB 251 or MICRB 251 or BIOL 230W or BMB 251H or BIOL 230M) or (MICRB 201 or MICRB 201H) Concurrent Courses: BMB 211 or BMB 401 or BMB 401H
Cross-Listed

BMB 443W: Laboratory in Protein Purification and Enzymology
3 Credits

This course is intended to provide advanced Biochemistry & Molecular Biology students with in class instruction on various laboratory methods used to explore the physical & chemical properties of proteins and other topics relevant to enzymes such as protein purification techniques and enzyme kinetic assays. In addition students will have hands-on laboratory experience conducting experiments related to 1) enzyme kinetics and product inhibition pattern to determine the order of product release, 2) purification of proteins such as rabbit muscle aldolase using a variety of protein purification methods including salt precipitation, dialysis, and ion-exchange/affinity chromatography using phosphocellulose resin, 3) determination of specific activity and purity of a student purified protein, 4) determination of molecular weight of native enzymes using gel-filtration-HPLC and subunit molecular weight by SDS-PAGE, 5) determination of isoelectric point and identification of C-terminal amino acid of a purified protein, and 6) determination of optimal conditions for protein crystal growth. Student performance is evaluated through a variety of assessments, including lab reports organized in accordance to the biochemistry journal articles and through written examinations that assess the understanding of principles and methodology.

Enforced Prerequisites: BMB 442 and (BMB 401 or BMB 401H)
Writing Across the Curriculum

BMB 445W: Laboratory in Molecular Genetics I
2 Credits

Laboratory in molecular techniques in gene analysis and microbial genetics, emphasizing in vitro methodologies. BMB 445W Laboratory in Molecular Genetics I (2) The objectives of BMB 445W are to provide advanced Biochemistry and Microbiology students with instruction in (1) techniques commonly used in modern research and clinical laboratories in this field, (2) evaluation of the quality of experimental data, including appropriate analysis, and (3) presentation of results of laboratory work in written form. Experiments are focused on key techniques and procedures such as DNA isolation, polymerase chain reaction, Southern hybridization analysis, cloning, and DNA sequencing. Students are evaluated via written lab reports and written examinations that assess the understanding of principles and methodology. BMB 445W is an extension of the nucleic acid section of BMB 442, which is a prerequisite for BMB 445W.
field of research. We have previously explored the general areas of cell biology, we delve into specific issues, often looking at "classic" publications exploring how innovative techniques have been applied to answer current questions in the context before progressing to the latest papers describing how a specific phenomenon was first investigated. Instead of a general survey of cell biology, we will read from journals to explore questions about cell biology and how these questions are addressed through scientific thinking as applied to experimental data and creative thinking about experimental approach exercises where students work in groups to practice posing new questions as suggested by our readings and proposing experiments to answer these questions. These skills are vital part of what cell biologists do daily, and these exercises provide practice in thinking like a scientist. Students have previously reported that by taking this course they acquired the ability to read and understand the primary literature and have gained an in-depth understanding about how to use various experimental techniques.

**Enforced Prerequisites:** BMB 442 and (MICRB 201 or MICRB 201H)

**Recommended Preparations:** (MICRB 202 or MICRB 203) and BMB 400

**BMB 448: Model Systems and Approaches in Cell Biology Inquiry**

2 Credits

Advanced laboratory that uses inquiry-based approaches to analyze organelles, genetic mechanisms, and metabolic processes in eukaryotic organisms.

**Enforced Prerequisites:** (BMB 251 OR MICRB 251 OR BIOL 230W OR BMB 251H OR BIOL 230M) AND (MICRB 202 OR MICRB 203) RECOMMENDED PREPARATIONS BMB 442

**BMB 450: Microbial/Molecular Genetics**

2 Credits

This course will focus on understanding the mechanisms and applications of the tools of both classical and molecular genetics to the study of prokaryotic organisms. To that end, topics to be discussed will include: 1) structure and replication of bacterial genomes (chromosomes, plasmids, etc.); 2) the organization of genes on bacterial genomes (operons, regulons, etc.); 3) regulation of gene expression; 4) mutagenesis and DNA repair; 5) recombination; 6) extrachromosomal DNA elements (plasmids, transposable elements and bacteriophages); 7) gene transfer in bacteria; 8) mechanisms of "immunity" to foreign DNA uptake; and 9) classical and molecular genetic analyses (complementation, conditional mutations, screens and selections, recombinant DNA, HTP techniques, nucleic acid mobility shift assays, blots, gene fusions, etc). The course will utilize a variety of formats including formal lecture presentations, guest lectures by BMB faculty performing research in the area of prokaryotic genetics, as well as student presentations.

**Enforced Prerequisites:** (BIOLOGY 222 OR BIOLOGY 322) AND (MICROBIOLOGY 201 OR MICROBIOLOGY 201H)

**Cross-listed with:** MICRB 450

**BMB 460: Cell Growth and Differentiation**

3 Credits

Mechanisms and regulation of protein trafficking, organelle biosynthesis, cell development, signaling and cell cycle control. Emphasizes experimental design and analysis. BMB 460 / MICRB 460 Cell Growth and Differentiation (3) is a unique course that uses the primary literature to teach significant content in advanced cell biology while simultaneously exposing students to the scientific craft of experimental design and analysis. In addition to exploring historical and current cell biology research articles, students will develop two vital scientific skills: critical thinking as applied to experimental data and creative thinking about solving unresolved questions in cell biology. In this course students will read from journals to explore questions about cell biology and how cell biologists decipher cell functions. Instead of a general survey of cell biology, we delve into specific issues, often looking at "classic" papers describing how a specific phenomenon was first investigated to place current questions in context before progressing to the latest publications exploring how innovative techniques have been applied to deciphering cell function. The course is divided into units, each of which emphasizes content in a different area. Actual content may vary from year to year as the course is updated to reflect progress in a field of research. We have previously explored the general areas of cell membrane dynamics, intracellular protein trafficking, cell cycle regulation, cell signaling pathways and cancer cell biology. Finally, the course ends with a unit on stem cells and therapeutic cloning technology. A portion of the final unit is also devoted to discussing the ethical implications of stem cell research with an emphasis on how to make personal decisions about how our society should approach these issues. Reading guides are provided for each assignment to help students find and understand important points in reading assignments. Class periods are devoted to explanations and instructor-led discussions about the readings with an emphasis on understanding the questions, the methods used to approach the questions, the experimental results, and the interpretations of the results. Furthermore, periodic class periods are dedicated to experimental approach exercises where students work in groups to practice posing new questions as suggested by our readings and proposing experiments to answer these questions. These skills are vital part of what cell biologists do daily, and these exercises provide practice in thinking like a scientist. Students have previously reported that by taking this course they acquired the ability to read and understand the primary literature and have gained an in-depth understanding about how to use various experimental techniques.

**Enforced Prerequisites:** BMB 251 OR MICRB 251 OR BMB 252H OR BIOL 230W OR BIOL 230M

**Cross-listed with:** MICRB 460

**BMB 460H: Cell Growth and Differentiation**

2 Credits

Mechanisms and regulation of protein trafficking, organelle biosynthesis, cell signaling, cell cycle control, and cell development.

**Cross-Listed Honors**

**BMB 464: Molecular Medicine**

3 Credits

An exploration of the impact of advances in molecular biology on understanding disease mechanisms, medical diagnosis, and therapeutics.

**Enforced Prerequisites:** BMB 251 OR MICRB 251 OR BIOL 230W OR BMB 251H OR BIOL 230M

**BMB 464H: Molecular Medicine Honors**

3 Credits

An exploration of the impact of advances in molecular biology on understanding disease mechanisms, medical diagnosis, and therapeutics.

**Enforced Prerequisites:** BMB 251 OR MICRB 251 OR BIOL 230W OR BMB 251H OR BIOL 230M

**BMB 465: Protein Structure and Function**

2 Credits

A study of the relationship between structure and function of proteins; internet analysis to predict structure and function is included.

**Enforced Prerequisite:** BIOL 230W
BMB 474: Analytical Biochemistry

3 Credits

Physical/chemical theory and techniques that emphasize purification and characterization of biological macromolecules, including proteins, lipids and nucleic acids. BMB 474 Analytical Biochemistry (3) This three-credit course deals with acquiring knowledge of laboratory skills required for success in experimental biochemistry and molecular biology. It is particularly suited for students intending a career as a research scientist in the areas of biochemistry, biotechnology, bioengineering, microbiology, or molecular biology. Course content focuses on the detection, purification and identification of biological macromolecules such as practice of separation science with emphasis on diffusion, gel permeation chromatography, ion-exchange chromatography, affinity chromatography, sedimentation velocity ultracentrifugation, sedimentation equilibrium ultracentrifugation, density gradient ultracentrifugation, agarose gel electrophoresis, SDS gel electrophoresis, isoelectrofocusing, membrane filtration and dialysis (including Donan equilibrium), ligand binding, high performance gas chromatography, high performance liquid chromatography, mass spectrometry, and immunological methods of macromolecules. The second unit includes the theory and practice of biological spectroscopy with emphasis on visible, infrared, circular dichroism, optical rotary dispersion, Raman, resonance Raman, nuclear magnetic resonance, electron paramagnetic resonance, Mossbauer, surface plasmon resonance, electron-nuclear double resonance, and electron spin echo spectroscopy of macromolecules. The lectures are designed to introduce a particular topic, to derive relevant equations, to supplement reading material with practical examples, and to clarify points in assigned problem sets. Two guest lectures by experts in the field will provide up-to-date information on mass spectroscopy and Mossbauer spectroscopy, and two site-visits, one to the mass spectrometry core facility and the other to the magnetic resonance core facility, will provide hands-on experience. Problem sets are not collected or graded; rather, answers are handed out in the following class period. This method provides the ability to collaborate with others on solving problems and to self-check work.

Enforced Prerequisite or concurrent: BMB 428 or CHEM 450

BMB 482: Introduction to Computational Biology

3 Credits

Modern DNA sequencing technologies have transformed molecular biology into a data science. Sequencing machines can now read hundreds of millions of DNA sequence fragments in a few hours and at low cost. These technologies not only enable affordable sequencing of individual genomes (human or any other species); they also allow us to investigate numerous ways in which the genome performs its biological functions in different cell types and how mutations in genomes give rise to various phenotypes. However, given the volume of data and the noisy nature of biological measurements and signals, we require intelligent and efficient computational algorithms to make sense of genomic datasets. The discipline of bioinformatics and computational biology aims to meet this need. This course focuses on understanding and applying the computational methods and algorithms that are used to analyze genomic data, in particular the large datasets arising from high-throughput DNA sequencing technologies. During the course, we will focus on several application areas in genomics that require computational analyses. These topics will be organized around three main themes: - Genomes: comparing DNA and protein sequences; locating sequences on the genome; assembling genomes. - Evolution: reconstructing evolutionary relationships; personal genomics; detecting disease-associated genome variations. - Function: understanding biochemical activities using functional genomics; discovering functional elements in genome sequences; characterizing regulatory relationships between genes. For each of the genomics topics listed above, we will focus on understanding the computational algorithms that are used to analyze data. Such algorithms may include dynamic programming (sequence alignment), graph algorithms (assembly), clustering methods (phylogenetics & metagenomics), and machine-learning approaches such as Expectation Maximization, Gibbs sampling, and Hidden Markov Models (various applications in discovering functional genomic elements). Students will also develop practical bioinformatics analysis skills throughout the course. Each bioinformatics topic will be accompanied by practical exercises. Students will also work in teams to research and develop a project that applies computational methods to a genomics-related problem.

Enforced Prerequisites: (BMB 252 OR MICRB 252 OR BMB 252H OR BIOL 230W OR BIOL 230M) AND (Biol 222 OR BIOL 322) RECOMMENDED PREPARATIONS MICRB 410 AND BMB 460 AND MICRB 415 Cross-listed with: MICRB 480
which are yielding exciting insights into answering these questions. One hallmark of genomic research is that data are released rapidly along with tools for browsing and analyzing it. Thus not only can you learn the major results by reading papers, but you can examine the underlying data and do your own analyses. Discovery is no longer the exclusive domain of the data producers - you can join in! This course will introduce students to ongoing research aimed at identifying functional regions in genomes and encourage them to use web-based bioinformatics tools for exploring the genomic and epigenetic data. Students will develop creative projects that address issues in functional genomics of high interest to them. The course has two phases, the first on the basics of genomics (sequencing, alignment, assembly, resources), and the second on the search for functional elements in genomes. The course will explore ways to find:- Protein-coding genes within genomes- Transcribed regions: How much of the genome is transcribed? Which transcribed regions do not code for proteins? What roles do they play in the cell (regulatory and enzymatic)?- Evolutionary signatures of function: How can you use genome comparisons between species to estimate the amount of functional sequence - and to identify it?- Non-genic functional sequences: How do you map epigenetic features associated with gene regulation, such as histone modifications, DNase hypersensitive sites, and transcription factor occupancy?- Function by phenotype: Given the ability of genetic association to find loci that contribute to complex traits, such as disease susceptibility, how does functional genomics aid in finding basis of these traits?

**Enforced Prerequisites:** (BMB 251 OR MICRB 251 OR BIOL 230W OR BMB 251H OR BIOL 230M) AND (BIOL 222 OR BIOL 322)

BMB 485: Human Genomics and Biomedical Informatics

3 Credits

This course covers the basics of measuring genomic variation and exploring how variation in DNA is related to common, complex disease.

**Enforced Prerequisite:** BIOL 322 and BMB 400 and (STAT 301 or STAT 401)

BMB 488: Communities of Practice in Biochemistry and Molecular Biology

2 Credits/Maximum of 16

The course combines laboratory research in a community of practice and a seminar on topics in science, ethics, and society.

**Enforced Prerequisites:** MICRB 202 RECOMMENDED PREPARATIONS
MICRB 202 OR MICRB 203

BMB 496: Independent Studies

1-18 Credits/Maximum of 18

Creative projects, including research and design, which are supervised on an individual basis and which fall outside the scope of formal courses.

BMB 497: Special Topics

1-9 Credits/Maximum of 9

Formal courses given infrequently to explore, in depth, a comparatively narrow subject which may be topical or of special interest.